

Article

Habitat and Season Effects on Small Mammal Bycatch in Live Trapping

Ines Hotopp¹, Bernd Walther² , Olaf Fuelling¹, Daniela Reil², Christin Hesse², Diana Alexandra Below², Christian Imholt² and Jens Jacob^{2,*}

¹ tier3 Solutions GmbH, Kolberger Str. 61–63, 51381 Leverkusen, Germany

² Rodent Research, Institute for Epidemiology and Pathogen Diagnostics, Julius Kühn-Institute (JKI)–Federal Research Centre for Cultivated Plants, Toppheideweg 88, 48161 Münster, Germany

* Correspondence: jens.jacob@julius-kuehn.de

Simple Summary: Trapping particular small mammal species is frequently used for scientific purposes but unnecessary bycatch can occur. Live trapping conducted over the last decade in Germany using Ugglan multiple capture traps in grassland, forest and margin habitats revealed about 30% bycatch when target species were common voles (*Microtus arvalis*) in grassland and common voles and bank voles (*Clethrionomys glareolus*) in margins and forests. This was more pronounced in spring and along margins. Species mentioned on the early warning list according to the Red List Germany were higher in numbers and proportion in spring and in grassland. The results of the study will help to avoid periods with enhanced presence of bycatch including endangered species (if the purpose of the study allows) or to pay particular attention in certain seasons and habitats when the occurrence of bycatch is most likely.

Abstract: Trapping small mammals is frequently used to study the dynamics, demography, behavior and presence of pathogens. When only particular small mammal species are in the focus of interest, all other species are unnecessary bycatch. We analyzed data from extensive live trapping campaigns conducted over the last decade in Germany, following a consistent standard trapping protocol that resulted in about 18,500 captures of small mammals. Animals were trapped with Ugglan multiple capture traps in grassland, forest and margin habitat. Trap success and the proportion of bycatch were about 30% when target species were common voles (*Microtus arvalis*) in grassland and common voles and bank voles (*Clethrionomys glareolus*) in margins and forests. This was more pronounced in spring and along margins. Species mentioned in the early warning list according to the Red List Germany were higher in numbers and proportion in spring and in grassland. The results will help to avoid periods with enhanced presence of bycatch, including endangered species (if the purpose of the study allows) or to pay particular attention in certain seasons and habitats when the occurrence of bycatch is most likely.

Keywords: conservation; non-target species; endangered species; rodents; Ugglan traps; voles



Citation: Hotopp, I.; Walther, B.; Fuelling, O.; Reil, D.; Hesse, C.; Below, D.A.; Imholt, C.; Jacob, J. Habitat and Season Effects on Small Mammal Bycatch in Live Trapping. *Biology* **2022**, *11*, 1806. <https://doi.org/10.3390/biology11121806>

Academic Editor: Carles Vilà

Received: 18 November 2022

Accepted: 7 December 2022

Published: 13 December 2022

Publisher's Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Copyright: © 2022 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

1. Introduction

Capturing wildlife is important to humans as wild animals are recognised as a source of nutrients and material for shelter, tools, medicine and cultural objects, etc. For millennia, wild animals have been vital in the process of domestication, for support in hunting and herding, lifting and transporting objects and other work, religious rites and as guards, as well as companions. The importance of wildlife in these aspects was later supplemented by the relevance of wildlife in research. In many cases, the species required needs to be obtained.

While capture for food does not necessarily require the animal to stay alive, their use for observational studies does. In some aspects of research such as the study of animal

populations, there is an obvious need not to disturb the object of study by unduly removing individuals [1]. In the last few decades, more and more emphasis has been put on the welfare of wildlife in research, including when trapping animals in the wild [2,3]. For research purposes, target animals should be placed in optimal conditions during the process of capturing, handling and releasing or transferring to research facilities, not only to ensure the welfare of animals [4] but also to ensure the validity of the research results obtained with them [5].

Rodents are the most species-rich mammalian order, with more than 2500 recent species [6]. Today, rodents are used for human [7,8] and animal food [9], pelts [10], entertainment [11], as pets [12,13] and, probably most importantly, for research [14,15]. Some rodent species are significant pests [16] and some can host and transmit zoonotic diseases to humans, pets and livestock [17]. Several rodent species are (critically) endangered or vulnerable [18].

Rodents are by far the vertebrate taxon most often bred and captured for research purposes. Wild rodents are obtained by live trapping and transferred to holding facilities (laboratories, enclosures) for various studies of their biology and ecology, including behavioural aspects [19,20] and risk assessment [21,22]. Others are trapped and released in situ for monitoring population dynamics [23,24], dispersal [25] or the epidemiology of rodent-borne pathogens [26,27].

Small mammals of a similar size to target species such as passerine birds, amphibians and reptiles are reported as bycatch in kill trapping [28,29] and live trapping [30,31]. Such bycatch can yield additional information as a rare, invasive and/or unseen/new species may be detected, but studies considering particular target species usually aim to minimize bycatch. While the death of animals is inevitable by design in kill traps, target and non-target animals may also die in live traps [32,33]. This can be due to fatal stress, bait that cannot be utilised by bycatch species or aggressive encounters between target and non-target animals caught in the same trap [34,35].

Attempts made to minimise bycatch include the choice of bait [36,37] and trap type most suitable for the target species [38,39], as well as optimal trapping location [40,41], optimal spacing [42,43] and signalling devices that allow for swift trap checks after capture [44]. In addition, modifications are made to reduce the accessibility of traps for non-target species such as a cover to exclude birds [45], trigger weight adjusted to the weight of the target species [46] or structures for escape [30] such as a “shrew hole” to let shrews escape [47,48].

Alternative methods to trapping that prevent bycatch include structures to sample hair [49,50] or scats [51], wildlife cameras [52] or indices of abundance that do not rely on trapping such as burrow counts [53], track counts [54,55], genetic methods based on non-invasive sampling methods [56] or chew cards and monitoring blocks [55,57].

However, in many circumstances, live trapping is the gold standard and often cannot be replaced by alternative methods. With live traps, often more individuals and more species can be captured than with snap traps [58,59]. Therefore, live trapping will remain the main method in studies of small mammals, be it for monitoring or the investigation of their population dynamics, demography or behaviour, while the study of associated pathogens often requires organ samples and the use of kill traps.

There is only limited knowledge through systematic and comprehensive long-term study of bycatch in live trapping. We analysed bycatch in live trapping data collected in Germany using Ugglan multiple capture traps over the last decade. Target species were the forest-dwelling bank vole (*Clethrionomys glareolus* syn. *Myodes glareolus*) and the grassland species common vole (*Microtus arvalis*). Both are small rodent species that are widely distributed across large parts of Europe [60] and regions in the western part of Asia [6]. The multi-annual dynamics are similar and characterised by population outbreaks every 2–5 years [61]. During outbreaks, common voles cause massive damage in agriculture and forestry throughout Europe [62], while large bank vole populations can cause damage in forestry [61] and increase the risk of transmission of Puumala orthohantavirus to humans [63,64]. For a better understanding of the patterns and processes related to out-

breaks, and to inform stakeholders in plant protection and health protection about the risks associated with outbreaks of these species, research and monitoring using live trapping are conducted regularly. Here, data from past trapping campaigns were re-analysed in order to gain a better understanding of patterns of the occurrence of bycatch. The effects of habitat, season and Red List status of non-target species were considered to identify when and where researchers should pay particular attention to minimise bycatch.

2. Materials and Methods

2.1. Field Sites

In four different studies, we used study sites totalling 28 forest, 8 grassland and 23 field margin habitats in six federal states of Germany (Table 1, Figure 1). All sites were at least 0.2–2.2 km apart from each other. Forest habitats were mainly beech forests or mixed deciduous forests dominated by beech (*Fagus sylvatica*), oak (*Quercus* spp.) and hornbeam (*Carpinus betulus*). Some forest sites were located at forest edges with dense understory vegetation. Grasslands were sown with perennial commercial seed mixes mainly used to produce fodder, including silage for dairy cattle or for grazing livestock. Margins were mostly hedgerows consisting of various woody species and shrubs that separated different arable fields or grasslands.

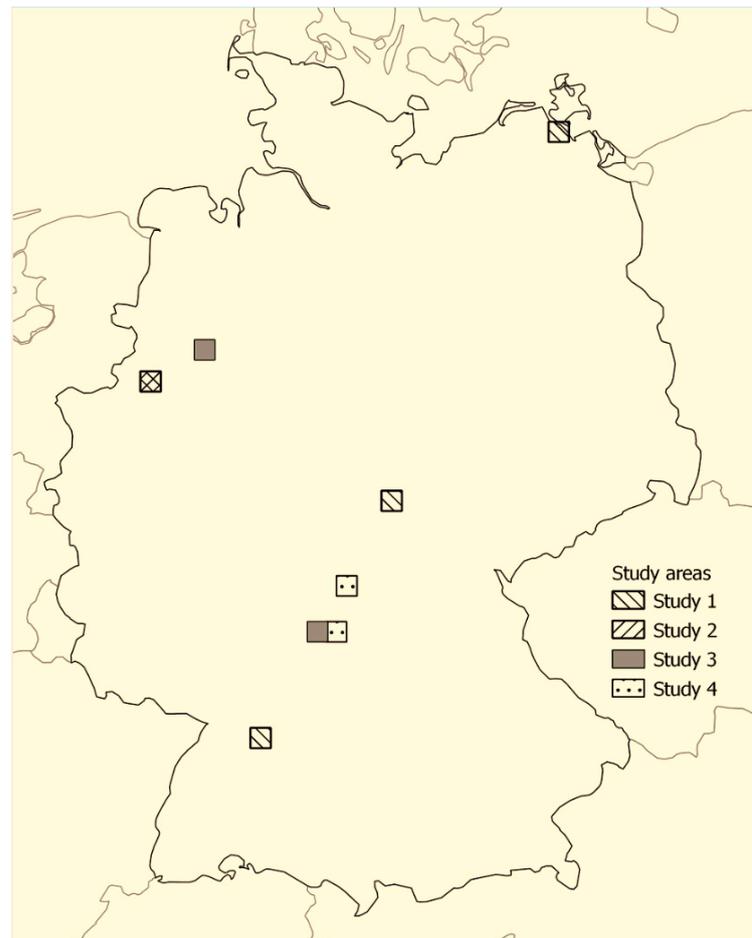


Figure 1. Location of study sites in Germany. © map shape: UIA World Countries Boundaries <https://hub.arcgis.com/maps/252471276c9941729543be8789e06e12> accessed 2 December 2022.

Table 1. Number of study sites, number of trap nights and number of captures of small mammals in Ugglan live traps from four studies conducted 2010–2021 in six federal states of Germany Baden-Wuerttemberg (BAW), Mecklenburg-Western Pomerania (MWP), North Rhine-Westphalia (NRW), Thuringia (THU), Bavaria (BAV), Lower Saxony (LOS).

Study	Years	Federal State	No. Study Sites				No. Trap Nights				No. Captures			
			Forest	Grassland	Margin	Total	Forest	Grassland	Margin	Total	Forest	Grassland	Margin	Total
# 1			16	8	2	26	39,788	11,172	4067	55,027	9851	1482	519	11,852
	2010–2014	BAW	4	3	0	7	9898	3675	0	13,573	2778	612	0	3390
	2010–2013	MWP	3	1	2	6	8330	2107	4067	14,503	2036	318	519	2873
	2010–2015	NRW	4	3	0	7	9506	3675	0	13,181	2074	181	0	2255
	2010–2013	THU	5	1	0	6	12,054	1715	0	13,769	2963	371	0	3334
# 2	2019–2021	NRW	12	0	0	12	27,500	0	0	27,500	5166	0	0	5166
# 3	2020	BAV	0	0	9	9	0	0	430	430	0	0	937	937
# 4			0	0	12	12	0	0	448.5	448.5	0	0	503	503
	2020	BAV	0	0	4	4	0	0	148.5	148.5	0	0	254	254
	2020	LOS	0	0	8	8	0	0	300	300	0	0	249	249
Total			28	8	23	59	67,538	8109	4945.5	83,355.5	15,017	1482	1959	18,458

2.2. Trapping

We used Ugglan live trap models No. 1 and No. 2 (Grahnbab, Gnosjö, Sweden) for trapping small mammals between March and November in 2010–2015 and 2019–2021. The Ugglan multi-capture live trap is frequently used in small mammal research all over Europe (e.g., [24,39]). The trap is made from wire mesh with a removable plastic floor for insulation and easy cleaning and an aluminium cover for weather protection. The trap is large enough to hold more than one small mammal and is usually equipped with bait and bedding. In studies 1 and 2 (Table 1), we arranged Ugglan live traps in grids of 7×7 (49) and 5×10 (50) traps with trap spacing of 7–10 m. In studies 3 and 4, we used 20–30 traps in 2–4 trap-lines 2–10 m apart and trap spacing of 10 m within lines. We pre-baited traps for 1–3 days before activating traps. In studies 1–2, bait was rolled oats supplemented with peanut curls, apple slices and rodent chow pellets and bedding was wood wool. In studies 3–4, bait was rolled oats, olive oil and dried mealworm supplemented with slices of apple, cucumber or carrot when temperatures were high and bedding when temperature was low.

In studies 1 and 2, live traps were set for three to four consecutive days and checked every 10–12 h, always around sunrise and sunset. In studies 3 and 4, we placed live traps for three consecutive nights, and set them each night for about six hours around sunset or sunrise. Within the six hours, traps were set and checked every 90–120 min. Animals were released at the point of capture. For the purpose of this analysis, we defined the bank vole and the common vole as the target species in forests and margins and the common vole as the target species in grassland. Both species are important model species in various scientific studies and in monitoring to assess the risk of rodent damage in forestry and agriculture or the risk of transmission of Puumala Orthohantavirus from bank voles to humans. However, the original studies might have targeted not just bank voles or common voles but other small mammal species as well. All other captured species were defined as non-target species for the present analysis.

2.3. Data Analyses

We analysed 18,458 captures of small mammals from 59 study sites obtained in 83,355.5 trap nights in nine years (Table 1). Only trapping occasions that yielded ≥ 5 captures were considered for analyses. We defined trap night as a period of approximately 12 h between trap checks independent of the time of the day (studies 1 and 2). A period of six hours between trap checks as in studies 3 and 4 was regarded as 0.5 trap nights.

The number of captures and the proportion per taxon, site and session was calculated as well as the number (captures per 100 trap nights [TN] [65] rounded to integers) and the proportion of non-target species in captures for each study site and trapping session. The Red List Germany (<https://www.rote-liste-zentrum.de/en/Download-Vertebrates-1874.html> accessed on 20 December 2021) [66] was used to allocate taxa according to Red List status. The proportion of captures of species on the early warning list (for simplicity labelled “endangered” throughout the manuscript) per site and session was calculated.

The effect of season and habitat on trap success and proportion of captures per taxon, non-target captures and red list status were assessed with program R, Vers. 4.1.1 [67]. Effects of the ecological factors habitat (forest, grassland, margin) and season (spring, summer, autumn) were analysed using generalised linear mixed models (GLMMs). Models were fitted by maximum likelihood estimation via Template Model Builder and Laplace approximation for the integration of random effects using the R-package glmmTMB, Vers. 1.1.2 [68]. If necessary, a zero-inflation model was included. Proportions were fitted with a binomial family and trap success was fitted using a negative binomial family to account for overdispersion in the count data. For each parameter, the model with the best fit according to the Akaike’s information criterion (AIC) [69] for fixed and random effect structures and, if applicable, zero-inflation formula was selected. The validity of the chosen models was checked by graphical evaluation of the normality of the residuals, the linearity between response and predictors, the independence of residuals and the calculation of the variance

inflation factors (VIFs). As the models contained interaction terms, VIFs could be expected to be high for some terms.

The final GLMMs for the proportion of non-target species included the explanatory variables habitat and season and their interaction. Year and study site nested per study were selected as random intercepts to account for repeated measurements and possible clustering of data along these factors. To assess the proportion of endangered species, the final GLMM included the interaction of all explanatory variables (habitat, season) and the random intercepts (year and study site). Figures were produced with the R-package ggplot2, Vers. 3.3.5 [70].

There were differences in the aim and design of studies 1–2 vs. 3–4 (see above). Studies 3–4 focussed on shrews in margins in spring and summer with frequent trap checks. This resulted in higher trap success, more Cricetidae and more non-target captures than in studies 1–2 that also covered seasons with low occurrence of Cricetidae. For the statistical analysis this is not ideal because even a simple calculation such as the mean per season can be skewed. An apparent temporal trend indicating low trap success in autumn is deceptive, because it is simply due to the missing data from studies 3–4. However, GLMMs are able to incorporate such misbalanced data. The study was included as a random effect and therefore the model includes the variability between the studies in calculations and predictions. Therefore, the results of the analysis of trap success per taxon and of non-target species captures are presented based on the model predictions and not as mean values. For the other variables, arithmetic means \pm standard errors are reported throughout. For all analyses, the level of statistical significance was set to $\alpha < 0.05$.

3. Results

During live trapping sessions in forest, grassland and margins, 18,458 small mammals of at least 15 species of four mammalian families were recorded (Tables 1 and 2). No other vertebrate species was trapped.

Trap success (captures per 100 TN) was highest in margins (40.0), followed by forests (22.3) and grasslands (13.3). In spring, the overall trap success was lowest (18.0) and it was highest in summer (26.7, autumn: 21.3) (Tables 2 and 3). In combination with habitat and season, trap success was highest in margins in spring (62.9) and lowest in grassland in spring (5.7) (Table 3).

On average, 34.1 (± 2.8) Cricetidae, 11.6 (± 2.0) Muridae and 3.0 (± 0.4) Soricidae were captured per 100 trap nights. The data contained many zeros and therefore the GLMM included a zero-inflation model. The model showed that Cricetidae were trapped twice as often as Muridae ($p < 0.05$) and 4.5 times as often as Soricidae ($p < 0.05$). Cricetidae were trapped 1.8 times more often in forests than in grasslands and margins ($p < 0.05$) (Figure 2a–c). For Muridae and Soricidae, trap success did not differ between habitats. According to model predictions, trap success of Cricetidae was 2.5 times higher in summer and in autumn 1.9 times higher than in spring (Figure 2a–c). In contrast, trap success of Muridae in summer was half as high as in spring ($p < 0.05$), but was similar in spring and autumn. Trap success of Soricidae in summer and autumn was at least 1.5 times higher than in spring ($p < 0.05$). Soricidae were more likely to be completely absent than the other two taxa ($p < 0.05$), and more likely to be completely absent in spring than in summer or autumn ($p < 0.05$).

The average proportion of Cricetidae was more than 5-fold higher than of Muridae and Soricidae ($p < 0.05$). It was highest in grassland and significantly lower in forests and margins (Figure 3a,b). A peak was reached in summer with a 1.3-fold increase ($p < 0.05$) compared to spring, but proportions were also significantly higher in autumn than in spring (1.2-fold increase) (Figure 3a,b). The proportion of Muridae was lowest in forests. In grassland, it was 1.4-fold increased ($p < 0.05$), and in margins 2.9-fold ($p < 0.05$) (Figure 3b). The proportion of Soricidae did not differ significantly among habitats, but was >15% lower in summer and autumn than in spring (Figure 3b). The likelihood of absence of Muridae and Soricidae was higher than for Cricetidae. For Soricidae it was lowest in spring ($p < 0.05$).

Table 2. Trap success (captures per 100 TN) and mean (per study site and session) of target and non-target small mammal taxa captured in Ugglan live traps in 28 forest, 8 grassland and 23 margin habitats from four different studies in Germany. Target species in forests and margins were the bank vole (*Clethrionomys glareolus*) and the common vole (*Microtus arvalis*) and in grasslands the common vole (*Microtus arvalis*). No other vertebrates than small mammals were captured, individuals that could not be allocated to a species were not considered further. SE—standard error. * Endangered species according to Red List Germany [66].

Taxon Species	All Habitats		Forest		Grassland		Margin	
	N 100 TN	Mean (±SE)	N 100 TN	Mean (±SE)	N 100 TN	Mean (±SE)	N 100 TN	Mean (±SE)
Cricetidae	16.65	32.38 (±0.14)	17.00	17.21 (±0.05)	11.26	12.11 (±0.40)	24.04	90.46 (±1.09)
Bank vole (<i>Clethrionomys glareolus</i>)	13.66	29.46 (±0.15)	15.81	16.59 (±0.06)	0.33	1.68 (±0.28)	14.50	80.50 (±1.24)
Common vole (<i>Microtus arvalis</i>)	2.82	23.71 (±0.3)	1.09	13.12 (±0.48)	10.56	12.86 (±0.45)	8.91	35.03 (±0.79)
Field vole (<i>Microtus agrestis</i>)	0.15	2.27 (±0.12)	0.09	1.61 (±0.07)	0.3	4.63 (±2.32)	0.63	3.16 (±0.77)
<i>Microtus</i> spp.	0.01	0.75 (±0.11)	0.01	0.55 (±0.08)	0.06	0.95 (±0.31)	0	0
Edible dormouse (<i>Glis glis</i>)	<0.01	0.41	0.00	0.41	0	0	0	0
Muridae	2.62	21.61 (±0.25)	2.2	4.65 (±0.04)	1.08	4.49 (±0.89)	11.97	59.12 (±1.26)
Yellow-necked mouse (<i>Apodemus flavicollis</i>)	1.77	21.46 (±0.31)	1.59	4.24 (±0.05)	0.07	1.09 (±0.39)	8.09	54.81 (±1.35)
Long-tailed field mouse (<i>Apodemus sylvaticus</i>)	0.25	11.9 (±0.33)	0.18	1.58 (±0.05)	0.08	1.84 (±1.01)	1.49	23.72 (±0.85)
Striped field mouse (<i>Apodemus agrarius</i>)	0.24	3.97 (±0.21)	0.14	2.64 (±0.24)	0	0	2.17	7.3 (±0.86)
<i>Apodemus</i> spp.	0.22	2.35 (±0.08)	0.26	2.24 (±0.07)	0.04	0.55 (±0.08)	0.02	11.11
Eurasian harvest mouse * (<i>Micromys minutus</i>)	0.14	3.24 (±0.51)	0.01	0.68 (±0.08)	0.90	6.80 (±1.94)	0.18	1.23 (±0.36)
House mouse (<i>Mus musculus</i>)	<0.01	1.22	<0.01	1.22	0	0	0	0
House rat (<i>Rattus rattus</i>)	<0.01	6.67	0	0	0	0	0.02	6.67
Mustelinae	0.02	0.53 (±0.04)	0.02	0.55 (±0.05)	0	0	0.02	0.41
Least weasel (<i>Mustela nivalis</i>)								
Soricidae	1.19	6.65 (±0.06)	1.03	2.88 (±0.03)	0.92	2.64 (±0.33)	3.98	14.46 (±0.26)
Common shrew (<i>Sorex araneus</i>)	0.05	19.14 (±0.93)	0	0	0	0	0.92	19.14 (±0.93)
Eurasian pygmy shrew (<i>Sorex minutus</i>)	0.03	10.26 (±0.34)	0	0.41	0	0	0.41	10.92 (±0.32)
<i>Sorex</i> spp.	1.08	2.88 (±0.03)	1.03	2.87 (±0.03)	0.92	2.64 (±0.33)	2.10	3.15 (±0.17)
Greater white-toothed shrew (<i>Crocidura russula</i>)	0.03	17.33 (±0.59)	0	0	0	0	0.49	17.33 (±0.59)
Eurasian water shrew * (<i>Neomys fodiens</i>)	0.01	5.05 (±0.81)	< 0.01	0.62 (±0.14)	0	0	0.06	8.00 (±0.00)
Not determined	1.67	6.79 (±0.10)	2.07	6.79 (±0.10)	0	0	0	0
All species	22.14	46.42 (±0.20)	22.32	22.05 (±0.06)	13.27	13.31 (±0.38)	40.02	142.05 (±1.51)

Table 3. Overall trap success per season (captures per 100 TN) and mean (per study site and session) of target and non-target small mammal taxa captured in Ugglan live traps in 28 forest, 8 grassland and 23 margin habitats from four different studies in Germany.

Taxon	Season	All Habitats		Forest		Grassland		Margins	
		N 100 TN	Mean (\pm SE)	N 100 TN	Mean (\pm SE)	N 100 TN	Mean (\pm SE)	N 100 TN	Mean (\pm SE)
Cricetidae	Spring	11.20	36.79 (\pm 0.49)	10.78	10.87 (\pm 0.15)	1.94	2.12 (\pm 0.18)	33.05	97.94 (\pm 2.24)
	Summer	22.25	40.91 (\pm 0.41)	22.94	22.68 (\pm 0.17)	16.20	17.28 (\pm 1.3)	26.49	95.34 (\pm 2.52)
	Autumn	15.93	16.50 (\pm 0.12)	16.8	17.44 (\pm 0.14)	13.13	14.01 (\pm 1.07)	9.84	9.27 (\pm 0.84)
Muridae	Spring	3.88	45.73 (\pm 0.97)	2.37	4.79 (\pm 0.13)	3.64	10.92 (\pm 3.8)	23.44	99.58 (\pm 2.69)
	Summer	1.95	7.77 (\pm 0.14)	2.03	4.38 (\pm 0.11)	0.05	0.41 (\pm 0.00)	4.98	14.61 (\pm 0.5)
	Autumn	2.17	4.71 (\pm 0.10)	2.20	4.78 (\pm 0.13)	0.28	0.98 (\pm 0.17)	7.65	7.23 (\pm 0.93)
Mustelinae	Spring	<0.01	0.41	<0.01	0.41	0	0	0	0
	Summer	0.01	0.41 (\pm 0.00)	0.02	0.41 (\pm 0.00)	0	0	0	0
	Autumn	0.03	0.65 (\pm 0.11)	0.03	0.72 (\pm 0.15)	0	0	0.07	0.41
Soricidae	Spring	0.94	11.51 (\pm 0.28)	0.62	2.31 (\pm 0.11)	0.10	1.22	6.40	18.91 (\pm 0.58)
	Summer	1.42	6.18 (\pm 0.12)	1.26	3.43 (\pm 0.13)	1.81	4.83 (\pm 1.39)	2.62	11.27 (\pm 0.43)
	Autumn	1.19	2.55 (\pm 0.04)	1.19	2.76 (\pm 0.05)	0.67	1.35 (\pm 0.15)	2.92	2.86 (\pm 0.35)
Incognita	Spring	2.02	10.34 (\pm 0.55)	2.46	10.34 (\pm 0.55)	0	0	0	0
	Summer	1.05	4.62 (\pm 0.23)	1.31	4.62 (\pm 0.23)	0	0	0	0
	Autumn	1.98	6.38 (\pm 0.21)	2.46	6.38 (\pm 0.21)	0	0	0	0
All species	Spring	18.03	67.25 (\pm 0.81)	16.23	15.80 (\pm 0.18)	5.68	5.68 (\pm 0.79)	62.90	191.92 (\pm 3.43)
	Summer	26.68	47.34 (\pm 0.44)	27.55	27.32 (\pm 0.19)	18.06	18.06 (\pm 1.26)	34.09	107.61 (\pm 2.58)
	Autumn	21.29	21.16 (\pm 0.14)	22.67	22.59 (\pm 0.18)	14.08	14.17 (\pm 0.99)	20.48	19.43 (\pm 1.39)

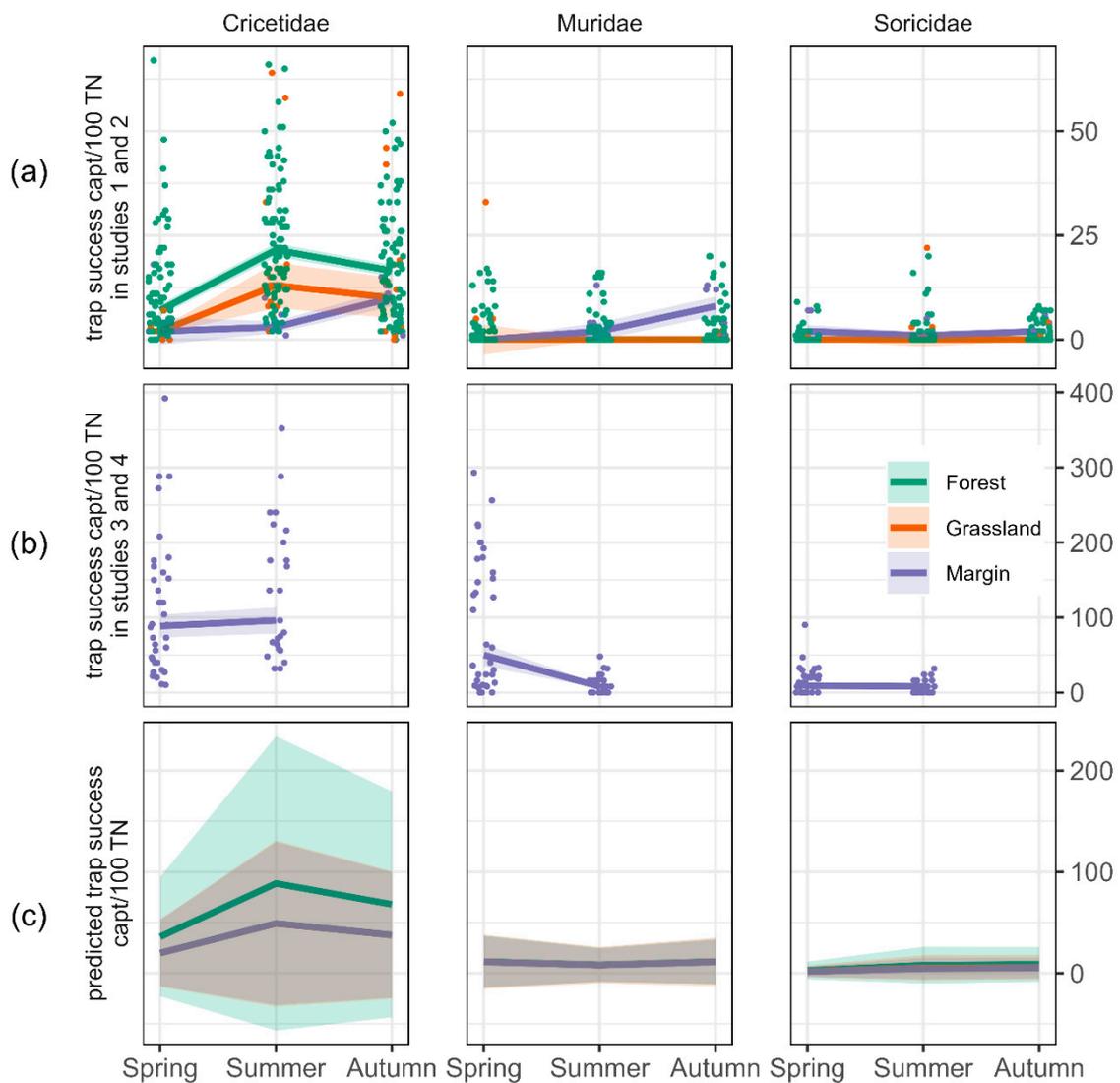


Figure 2. (a) Mean trap success (captures per 100 trap nights) per taxon, habitat and season in studies 1 and 2, (b) mean trap success (captures per 100 trap nights) per taxon, habitat and season in studies 3 and 4. Coloured areas in (a,b) depict the standard error. Means are based on multiple sites and trapping occasions (coloured dots). Means are presented separately for studies 1–2 and 3–4 to account for the otherwise skewed presentation due to varying study designs. (c) Predicted trap success by a GLMM with negative binomial family. Coloured areas in (c) depict the confidence interval of the statistical model. Note the different scales of the y-axes.

Trap success of non-target species (16.4 captures per 100 TN ± 2.1) was on average approximately half of the trap success of target species (34.0 captures per 100 TN ± 2.9). Model predictions suggested similar trap success of non-target species for margins, forest and grassland and for seasons (Figure 4a–c). The absence of non-target species was more likely in grassland than in forests and in spring than in autumn ($p < 0.05$) (Figure 4a–c).

The average proportion of non-target species captures was 0.71 (± 0.01). The proportion of non-target species captures was 50% higher in margins than in forest ($p < 0.05$) but did not differ between grassland and forest (Figure 5a,b). It was 100% higher in spring than in summer ($p < 0.05$) and 30% lower in autumn than in spring (Figure 5a,b) ($p < 0.05$).

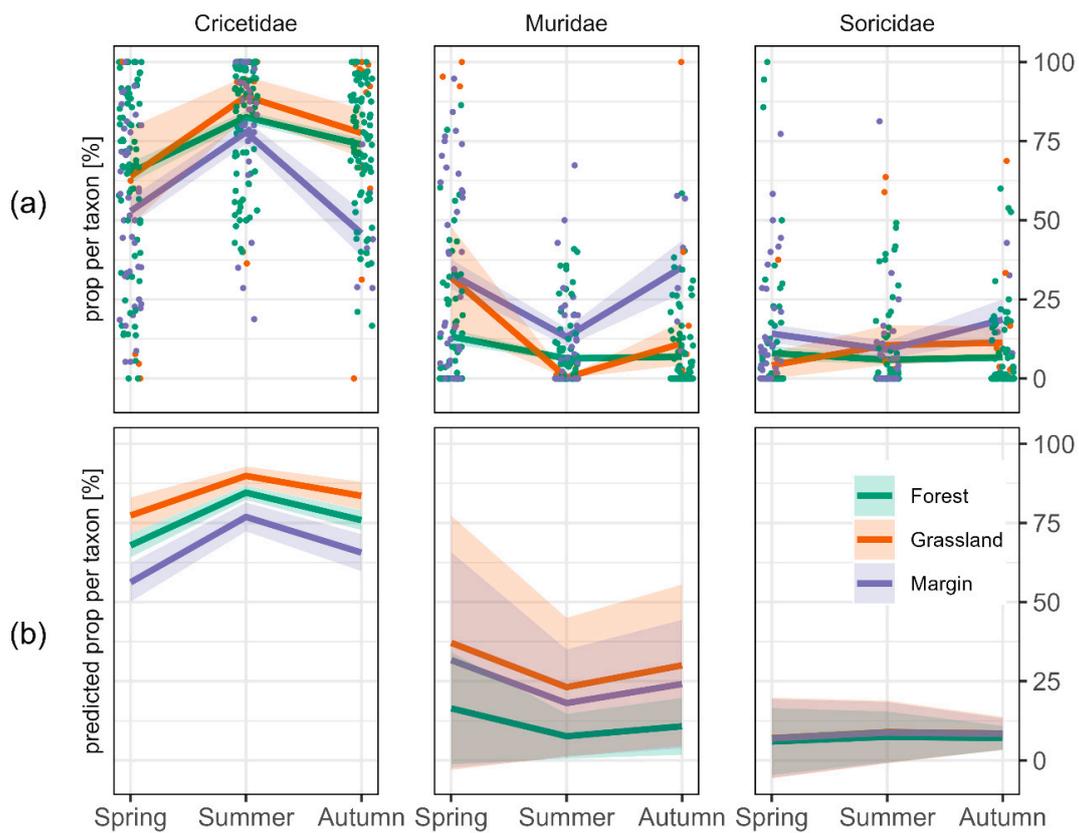


Figure 3. (a) Mean values for the proportion of captures per taxon, habitat and season. Coloured areas depict the standard error. Means are based on multiple sites and trapping occasions (coloured dots), and (b) predicted proportions by a GLMM with binomial family. Coloured areas in (b) depict the confidence interval of the statistical model.

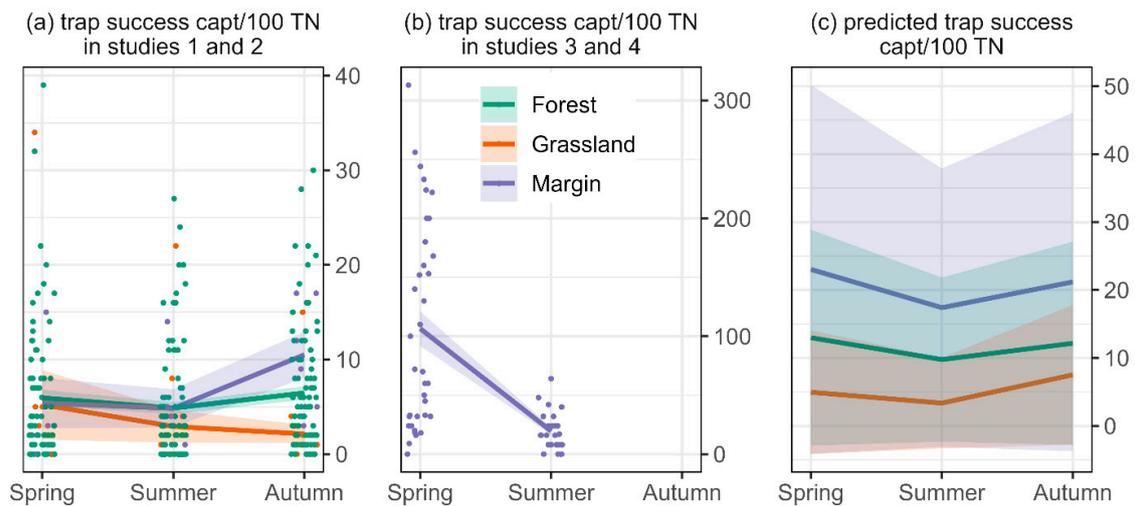


Figure 4. (a) Mean trap success of non-target captures (captures per 100 trap nights) per habitat and season in studies 1 and 2, (b) mean trap success of non-target captures (captures per 100 trap nights) per habitat and season in studies 3 and 4. Coloured areas in (a,b) depict the standard error. Means are based on multiple sites and trapping occasions (coloured dots). Means are presented separately for studies 1–2 and 3–4 to account for the otherwise skewed presentation due to different study designs. (c) Predicted trap success with a GLMM with negative binomial family. Coloured areas in (c) depict the confidence interval of the statistical model. Note the different scales of the y-axes.

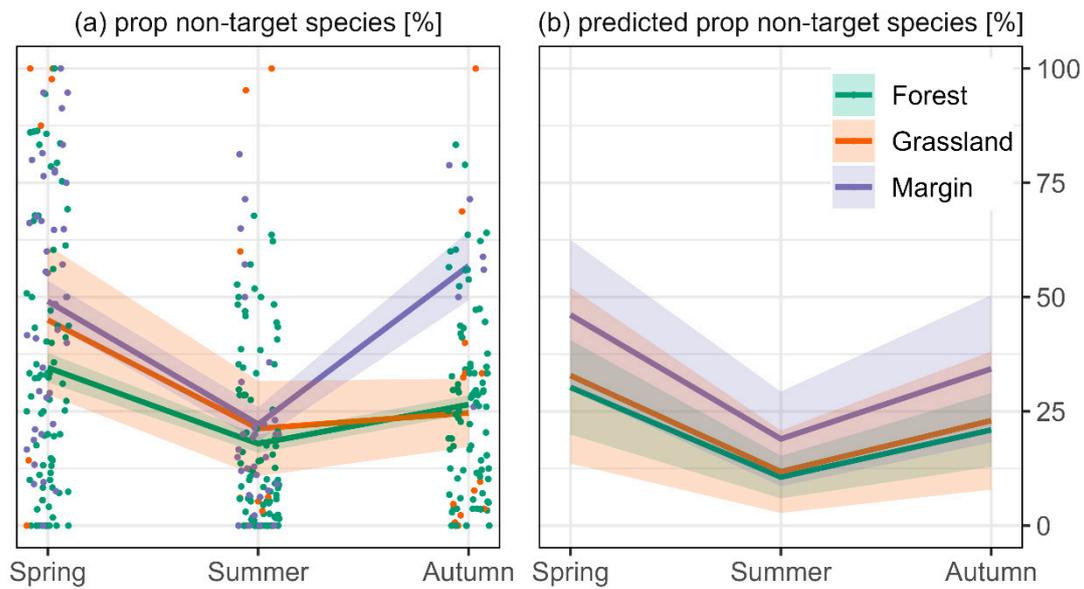


Figure 5. (a) Mean proportion of non-target captures per habitat and season. Coloured areas depict the standard error. Means are based on multiple sites and trapping occasions (coloured dots), and (b) predicted proportion of non-target species by GLMM with binomial family. Coloured areas in (b) depict the confidence interval of the statistical model.

The occurrence of endangered species (*Micromys minutus* and *Neomys fodiens*) trapped in the studies was too scarce for a thorough statistical analysis. Available data suggest an increased presence of endangered species in grassland compared to margins (3.2-fold increase) and forests (54-fold increase) (Figure 6a) and higher trap success in spring compared to summer (1.75-fold increase) and autumn (4.3-fold increase) (Figure 6a). Similarly, the proportion of endangered species seemed highest in grassland and in spring (Figure 6b).

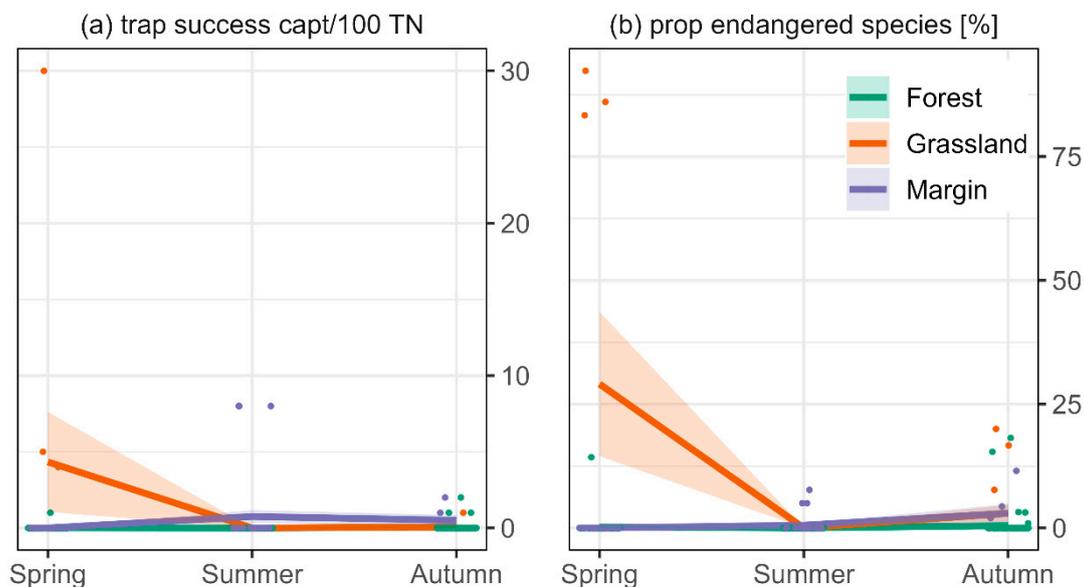


Figure 6. (a) Mean number (captures per 100 trap nights) and (b) mean proportion of endangered species (according to Red List Germany [66]) that were captured per habitat and season. Coloured areas in (b) depict the standard error. Means are based on multiple sites and trapping occasions (coloured dots). Note the different scales of the y-axes.

4. Discussion

To our knowledge, this is the first attempt to investigate live trapping data from central Europe regarding the distribution of captures of target and non-target taxa across habitats, Red List status and season. The study shows that, overall, about a third of live trapped small mammals were bycatch. This is not surprising because the small mammal composition of central European deciduous forests comprises a variety of small mammal species [71] dominated by bank voles and *Apodemus* species [72,73]. The same is true for open landscape habitats adjacent to hedgerows, forests and human settlements, where common voles are most prevalent [74,75].

The considerable proportion of non-target animals in standard live trapping is unwanted but less worrying than in kill trapping because animals can be released. However, there can be mortality in small mammals trapped alive [32,33]. Bycatch is also known from trapping programs of other taxa such as invertebrates [76,77] and birds [78]. Insect bycatch can be reduced using species-specific pheromones as lures [79] and specific birds such as starlings (*Sturnus vulgaris*) can be attracted by placing a conspecific individual close to the trapping device or using species-specific bird calls [80]. Species-specificity cannot be achieved with Uggan traps considered here or with other standard live traps used in small mammal research such as Longworth traps or Sherman traps because they all, by design, capture all animals small enough to pass through the trap's entrance [39,81].

Generally, the trap success of non-target taxa was higher in forest versus margins and grassland and higher in spring than summer or autumn. Naturally, the more restricted the number of target species, the higher the proportion of non-target species trapped. We have used an extreme approach focussing on only 1–2 target species in each habitat, albeit rather common ones. Therefore, the proportion of non-target species we report is likely to be drastically reduced when several species are targets.

Habitat mattered for bycatch because trap success of non-target species was higher in margins than in forest habitats. This finding seems plausible as in central Europe, open agricultural habitats including grassland that stretch far between margins are of poor small mammal diversity. Grassland tends to host the common vole as the main species with few other small mammals present [75,82]. The woody plants and shrubs in margins provide more complex horizontal and vertical structures that can be a suitable habitat or provide a dispersal corridor for several small mammal species [71,83]. Non-targets were obviously attracted to the live traps at habitat interfaces, leading to an elevated proportion in margins.

Seasonally, the proportion of non-target taxa was highest in spring independent of habitat. Several factors may have caused this result. Spring temperatures are lower than in summer and autumn [84], body condition after winter tends to be low [85,86] and age of animals high [87], which may all cause increased need for food uptake. However, in spring, food availability is low. This could have caused non-target species more than target species to utilise bait in traps more intensely than they might do in other seasons. Such species may also tolerate the (previous) presence of other species in and around traps in spring more than in other seasons [88,89]. Additionally, limited food supply in spring may prompt non-target species to accept potential predation risk associated with increasing home ranges and approaching traps in an otherwise less preferred landscape of risk [90,91]. All this could result in increased captures of non-target species but this needs to be validated in future work.

We used data from previous studies that were not designed a priori to assess bycatch and therefore were slightly different in bait used, trapping schedule and trap type. However, the large numbers of trap nights and captures yielded a robust indication where main non-target issues can occur. It seems unrealistic to run similarly large live trapping campaigns solely dedicated to an assessment of bycatch but there may be the opportunity to include information from further studies (other trap types, habitats, species) in the future.

Most bycatch consisted of taxa not considered endangered. This reflects that endangered species are rare and therefore rarely encountered. There were two species (Eurasian water shrew, Eurasian harvest mouse) trapped that have an early warning status according

to the Red List Germany (<https://www.rote-liste-zentrum.de/en/Download-Vertebrates-1874.html> accessed on 20 December 2021) [66]. The trap success of Eurasian water shrews was the lowest of all species (0.0001 captures per 100 TN) and Eurasian harvest mice occurred in <4% of trapping sessions. Similar to all non-target captures, the seasonal dynamics of the proportion of endangered non-target taxa seemed highest in spring and higher in forests. This indicates that endangered species follow the same capture patterns and seem to utilise traps similar to all non-target species.

There are techniques available to minimise bycatch including bait that is attractive to the target and unattractive to the non-target species. However, many rodent species seem to prefer similar bait or similar bait properties [92,93], which makes it difficult to use a species-specific attractant, but see [36,94]. Trap placement on runways or close to the burrows of target species might limit bycatch but interferes with standardised trapping grids or transect designs. The size of the trap entrance can be minimised to reflect the body size of the target species to exclude all larger taxa. Trap placement away from instead of along margins/habitat interfaces is likely to minimise bycatch but the purpose of the study may require trapping in such locations. Trap sensors are available to indicate capture and they can be used to allow swift removal of trapped animals [44] instead of relying on a fixed trap check frequency. Any method to minimise bycatch needs to be evaluated carefully to make sure that there is no interference with the purpose of the study to guarantee valid result; this is paramount for scientific work. In addition, while maintaining the desired level of animal protection, methods need to be feasible economically regarding workload. This balance might be difficult to find.

5. Conclusions

The result presented here can be used to apply protective measures in a targeted fashion and to exclude periods when captures of non-targets and especially endangered species are less probable. Particular attention needs to be paid when trapping in spring along margins if all non-target taxa are to be avoided. For minimising the number and proportion of endangered species, trapping in grassland in spring seems most critical. These times and periods are likely to change when other species are targeted than in this study.

Further parameters potentially related to the risk of trapping non-target species and the risk of mortality of small mammals in traps include temperature, day/night activity and inter-specific competition. Comprehensive data from previous studies should be available to assess such effects and results could be used to further fine-tune measures to minimise the number, proportion and mortality of non-target taxa in live trapping.

Author Contributions: Conceptualization, J.J. and B.W.; methodology, J.J., B.W., I.H. and O.F.; formal analysis, I.H. and B.W.; investigation, I.H., B.W., O.F., D.R., C.H., D.A.B., C.I. and J.J.; data curation, I.H. and B.W.; writing—original draft preparation, J.J., B.W., I.H. and O.F.; writing—review and editing, I.H., B.W., O.F., D.R., C.H., D.A.B., C.I. and J.J.; visualization, I.H.; supervision, J.J. All authors have read and agreed to the published version of the manuscript.

Funding: Live trapping data originated, inter alia, from studies commissioned and funded by the Federal Environment Agency within the Environment Research Plan of the German Federal Ministry for the Environment, Nature Conservation and Nuclear Safety (BMU) (grant #370941401, #371348401) and the federal Ministry of Education and Research (BMBF) (grant #01KI1721E, #01KI2004E). Some of the live trapping data from summer 2020 originate from fieldwork funded by commercial sponsors and were provided with their kind permission.

Institutional Review Board Statement: Regierung von Unterfranken #55.2.2-2532.2.1062-15, 23.01.2020, #55.2.2-2532-2-1062-20 26.02.2020; Regierungspräsidium Stuttgart #35-9185.82/0261 09.04.2010; Landesamt für Natur, Umwelt und Verbraucherschutz Nordrhein-Westfalen #8.87-51.05.20.09.210 26.10.2009, #84-02.04.2014.A227 04.09.2014, #81-02.04.2018.A150 18.01.2019; Niedersächsisches Landesamt für Verbraucherschutz und Lebensmittelsicherheit Landesamt für Landwirtschaft #33.19-42502-04-20/3412 (09.06.2020); Landesamt für Lebensmittelsicherheit und Fischerei Mecklenburg-Vorpommern #7221.3-030/09 04.11.2009; Thüringer Landesamt für Lebensmittelsicherheit und Verbraucherschutz #22-2684-04-15-107/09 07.12.2009.

Informed Consent Statement: Not applicable.

Data Availability Statement: Data supporting reported results can be obtained from the authors. Access to data from studies 3–4 is subject to in-confidence regulation with commercial sponsor.

Acknowledgments: We thank all landowners for allowing us to work on their properties and field staff for assistance.

Conflicts of Interest: The authors declare no conflict of interest. The funders had no role in the design of the study; in the collection, analyses, or interpretation of data; in the writing of the manuscript; or in the decision to publish the results.

References

1. Sullivan, T.P.; Sullivan, D.S.; Ransome, D.B.; Lindgren, P.M.F. Impact of removal-trapping on abundance and diversity attributes in small-mammal communities. *Wildl. Soc. Bull.* **2003**, *31*, 464–474.
2. Baker, S.; Shaw, R.; Atkinson, R.; West, P.; Macdonald, D. Potential welfare impacts of kill-trapping European moles (*Talpa europaea*) using scissor traps and Duffus traps: A post mortem examination study. *Anim. Welf.* **2015**, *24*, 1–14. [CrossRef]
3. European Union–Canada–Russian Federation. Agreement on international humane trapping standards between the European Community, Canada and the Russian Federation. *Off. J. Eur. Communities* **1998**, *L42*, 43–57.
4. Fall, M.W. The search for acceptable animal traps. In Proceedings of the 20th Vertebrate Pest Conference, Reno, NV, USA, 4–7 March 2002; pp. 371–377.
5. Stephens, R.B.; Anderson, E.M. Effects of trap type on small mammal richness, diversity, and mortality. *Wildl. Soc. Bull.* **2014**, *38*, 619–627. [CrossRef]
6. Burgin, C.J.; Colella, J.P.; Kahn, P.L.; Upham, N.S. How many species of mammals are there? *J. Mammal.* **2018**, *99*, 1–14. [CrossRef]
7. Gruber, K. Rodent meat—a sustainable way to feed the world?: Using rodents as food has a long tradition in many parts of the world. *EMBO Rep.* **2016**, *17*, 630–633. [CrossRef]
8. Assogbadjo, A.E.; Codjia, J.T.C.; Sinsin, B.; Ekue, M.R.M.; Mensah, G.A. Importance of rodents as a human food source in Benin. *Belg. J. Zool.* **2005**, *135*, 11–15.
9. Kerr, K.R.; Kappen, K.L.; Garner, L.M.; Swanson, K.S. Commercially available avian and mammalian whole prey diet items targeted for consumption by managed exotic and domestic pet felines: Macronutrient, mineral, and long-chain fatty acid composition. *Zoo Biol.* **2014**, *33*, 327–335. [CrossRef]
10. Strege, G. Fur as fashion in America. *Fash. Style Pop. Cult.* **2014**, *1*, 413–432. [CrossRef]
11. Hitchens, P.L.; Booth, R.H.; Stevens, K.; Murphy, A.; Jones, B.; Hemsworth, L.M. The welfare of animals in Australian filmed media. *Animals* **2021**, *11*, 1986. [CrossRef]
12. Brown, C.J.; Donnelly, T.M. Rodent husbandry and care. *Vet. Clin. Exot. Anim. Pract.* **2004**, *7*, 201–225. [CrossRef]
13. Grant, R.A.; Montrose, V.T.; Wills, A.P. ExNOTic: Should we be keeping exotic pets? *Animals* **2017**, *7*, 47. [CrossRef]
14. DFG. *Animal Experimentation in Research*; Deutsche Forschungsgemeinschaft (DFG, German Research Foundation): Bonn, Germany, 2016; p. 76.
15. Carbone, L. Estimating mouse and rat use in American laboratories by extrapolation from animal welfare act-regulated species. *Sci. Rep.* **2021**, *11*, 493. [CrossRef]
16. Buckle, A.P.; Smith, R.H. *Rodent Pests and Their Control*, 2nd ed.; CAB International: Wallingford, Oxon, UK, 2015; p. 432.
17. Battersby, S.A. Rodents as carriers of disease. In *Rodent Pests and Their Control*, 2nd ed.; CAB International: Wallingford, Oxon, UK, 2015; pp. 81–100.
18. IUCN. *The IUCN Red List of Threatened Species*; Version 2014.3; IUCN: Gland, Switzerland, 2014.
19. Jedrzejewska, B.; Jedrzejewski, W. Antipredatory behaviour of bank voles and prey choice of weasels—enclosure experiments. *Ann. Zool. Fenn.* **1990**, *27*, 321–328. Available online: <https://www.jstor.org/stable/23734571> (accessed on 20 December 2021).
20. Mazza, V.; Eccard, J.A.; Zaccaroni, M.; Jacob, J.; Dammhahn, M. The fast and the flexible: Cognitive style drives individual variation in cognition in a small mammal. *Anim. Behav.* **2018**, *137*, 119–132. [CrossRef]
21. Edge, W.D.; Schaubert, E.M. Factors affecting risk assessment of small mammals to pesticides. *Environ. Toxicol. Chem.* **2000**, *19*, 2735–2741. [CrossRef]
22. Sheffield, S.R.; Lochmiller, R.L. Effects of field exposure to diazinon on small mammals inhabiting a semienclosed prairie grassland ecosystem. I. Ecological and reproductive effects. *Environ. Toxicol. Chem.* **2001**, *20*, 284–296. [CrossRef]
23. Singleton, G.R. Population dynamics of an outbreak of house mice (*Mus domesticus*) in the mallee wheatlands of Australia—hypothesis of plague formation. *J. Zool.* **1989**, *219*, 495–515. [CrossRef]
24. Hein, S.; Jacob, J. Population recovery of a common vole population (*Microtus arvalis*) after population collapse. *Pest Manag. Sci.* **2018**, *75*, 908–914. [CrossRef]
25. Flowerdew, J.R.; Shore, R.F.; Poulton, S.M.; Sparks, T.H. Live trapping to monitor small mammals in Britain. *Mammal Rev.* **2004**, *34*, 31–50. [CrossRef]

26. Bagamian, K.H.; Towner, J.S.; Kuenzi, A.J.; Douglass, R.J.; Rollin, P.E.; Waller, L.A.; Mills, J.N. Transmission ecology of Sin Nombre hantavirus in naturally infected North American deer mouse populations in outdoor enclosures. *PLoS ONE* **2012**, *7*, e47731. [[CrossRef](#)] [[PubMed](#)]
27. Douglass, R.J.; Vadell, M.R. How much effort is required to accurately describe the complex ecology of a rodent-borne viral disease? *Ecosphere* **2016**, *7*, e01368. [[CrossRef](#)] [[PubMed](#)]
28. Peitz, D.G.; Tappe, P.A.; Thill, R.E.; Perry, R.W.; Melchior, M.A.; Wigley, T.B. Non-target captures during small mammal trapping with snap traps. *Proc. Annu. Conf. Southeast. Assoc. Fish Wildl. Agencies* **2001**, *55*, 382–388.
29. Walther, B.; Geduhn, A.; Schenke, D.; Jacob, J. Exposure of passerine birds to brodifacoum during management of Norway rats on farms. *Sci. Total Environ.* **2021**, *762*, 144160. [[CrossRef](#)]
30. Karraker, N.E. String theory: Reducing mortality of mammals in pitfall traps. *Wildl. Soc. Bull.* **2001**, *29*, 1158–1162. Available online: <https://www.jstor.org/stable/3784139> (accessed on 20 December 2021).
31. Cook, W.M. Inadvertent bird captures in Sherman small mammal traps in an old field mosaic. *Trans. Kans. Acad. Sci.* **2004**, *107*, 170–172. [[CrossRef](#)]
32. Montgomery, W.I. Mortality of small rodents captured in live-traps. *Acta Theriol.* **1980**, *25*, 277–294. [[CrossRef](#)]
33. Corke, D. The deaths of small mammals in live-traps. *J. Zool.* **1967**, *153*, 552.
34. Kalinowska, A. Trapping of *Apodemus flavicollis* and *Clethrionomys glareolus* into a double trap. *Acta Theriol.* **1971**, *16*, 73–78. [[CrossRef](#)]
35. Waudby, H.P.; Petit, S.; Gill, M.J. The scientific, financial and ethical implications of three common wildlife-trapping designs. *Wildl. Res.* **2019**, *46*, 690–700. [[CrossRef](#)]
36. Schlötterburg, A.; Jakob, G.; Bellingrath-Kimura, S.; Jacob, J. Natural bait additives improve trapping success of common voles, *Microtus arvalis*. *Appl. Anim. Behav. Sci.* **2018**, *208*, 75–81. [[CrossRef](#)]
37. Takács, S.; Musso, A.E.; Gries, R.; Rozenberg, E.; Borden, J.H.; Brodie, B.; Gries, G. New food baits for trapping house mice, black rats and brown rats. *Appl. Anim. Behav. Sci.* **2017**, *200*, 130–135. [[CrossRef](#)]
38. Jacob, J.; Ylönen, H.; Hodgkinson, C.G. Trapping efficiency of Ugglan traps and Longworth traps for house mice in southeastern Australia. *Wildl. Res.* **2002**, *29*, 101–103. [[CrossRef](#)]
39. Jung, T.S. Comparative efficacy of Longworth, Sherman, and Ugglan live-traps for capturing small mammals in the Nearctic boreal forest. *Mammal Res.* **2015**, *61*, 57–64. [[CrossRef](#)]
40. Cunningham, R.B.; Lindenmayer, D.B.; MacGregor, C.; Barry, S.; Welsh, A. Effects of trap position, trap history, microhabitat and season on capture probabilities of small mammals in a wet eucalypt forest. *Wildl. Res.* **2005**, *32*, 657–671. [[CrossRef](#)]
41. Weihong, J.; Veitch, C.R.; Craig, J.L. An evaluation of the efficiency of rodent trapping methods: The effect of trap arrangement, cover type, and bait. *N. Zealand J. Ecol.* **1999**, *23*, 45–51.
42. Tew, T.E.; Todd, I.A.; MacDonald, D.W. The effects of trap spacing on population estimation of small mammals. *J. Zool.* **1994**, *233*, 340–344. [[CrossRef](#)]
43. Bovendorp, R.S.; McCleery, R.A.; Galetti, M. Optimising sampling methods for small mammal communities in neotropical rainforests. *Mammal Rev.* **2017**, *47*, 148–158. [[CrossRef](#)]
44. Notz, E.; Imholt, C.; Reil, D.; Jacob, J. Testing automated sensor traps for mammal field studies. *Wildl. Res.* **2017**, *44*, 72–77. [[CrossRef](#)]
45. Lane, V.R.; Miller, K.V.; Castleberry, S.B.; Miller, D.A.; Wigley, T.B. Methods to reduce avian bycatch in small mammal studies using snap traps. *J. Wildl. Manag.* **2010**, *74*, 595–599. [[CrossRef](#)]
46. Gurnell, J.; Flowerdew, J.R. Live trapping small mammals. A practical guide. *Publ. Mammal Soc.* **1982**, *Occasional Publication no. 3*, 1–24.
47. Little, J.; Gurnell, J. Shrew Captures and Rodent Field Studies. *J. Zool.* **1989**, *218*, 329–331. [[CrossRef](#)]
48. Eccard, J.A.; Klemme, I. Reducing mortality of shrews in rodent live trapping—a method increasing live-trap selectivity with shrew exits. *Ann. Zool. Fenn.* **2013**, *50*, 371–376. [[CrossRef](#)]
49. Chiron, F.; Hein, S.; Chargé, R.; Julliard, R.; Martin, L.; Roguet, A.; Jacob, J. Validation of hair tubes for small mammal population studies. *J. Mammal.* **2018**, *99*, 478–485. [[CrossRef](#)]
50. Barja, I.; Navarro-Castilla, Á.; Pérez, L. Effectiveness and applications of hair traps for the study of wild mammal populations. *Pol. J. Ecol.* **2016**, *64*, 409–419. [[CrossRef](#)]
51. Tennant, E.N.; Cypher, B.L.; Saslaw, L.R.; Westall, T.L.; Mohay, J.L.; Kelly, E.C.; Job, C.L.V. Evaluation of noninvasive survey methods for detecting endangered shrews. *Wildl. Soc. Bull.* **2020**, *44*, 610–616. [[CrossRef](#)]
52. Rendall, A.R.; Sutherland, D.R.; Cooke, R.; White, J. Camera trapping: A contemporary approach to monitoring invasive rodents in high conservation priority ecosystems. *PLoS ONE* **2014**, *9*, e86592. [[CrossRef](#)]
53. Fauteux, D.; Gauthier, G.; Mazerolle, M.J.; Coallier, N.; Bety, J.; Berteaux, D. Evaluation of invasive and non-invasive methods to monitor rodent abundance in the arctic. *Ecosphere* **2018**, *9*, e02124. [[CrossRef](#)]
54. Carey, A.B.; Witt, J.W. Track counts as indexes to abundances of arboreal rodents. *J. Mammal.* **1991**, *72*, 192–194. [[CrossRef](#)]
55. Whisson, D.A.; Engeman, R.M.; Collins, K. Developing relative abundance techniques (RATs) for monitoring rodent populations. *Wildl. Res.* **2005**, *32*, 239–244. [[CrossRef](#)]

56. Ferreira, C.M.; Sabino-Marques, H.; Barbosa, S.; Costa, P.; Encarnacao, C.; Alpizar-Jara, R.; Pita, R.; Beja, P.; Mira, A.; Searle, J.B.; et al. Genetic non-invasive sampling (gNIS) as a cost-effective tool for monitoring elusive small mammals. *Eur. J. Wildl. Res.* **2018**, *64*, 64. [[CrossRef](#)]
57. Sweetapple, P. Chew-track-cards: A multiple-species small mammal detection device. *N. Zealand J. Ecol.* **2011**, *35*, 153–162.
58. Hansson, L.; Hoffmeyer, I. Snap and live trap efficiency for small mammals. *Oikos* **1973**, *24*, 477–478. [[CrossRef](#)]
59. Lee, L.-L. Effectiveness of live traps and snap traps in trapping small mammals in kinmen. *Acta Zool. Taiwanica* **1997**, *8*, 79–85.
60. Niethammer, J.; Krapp, F. *Handbuch der Säugetiere Europas Band 2/I Nagetiere II*; Akademische Verlagsgesellschaft: Wiesbaden, Germany, 1982.
61. Jacob, J.; Tkadlec, E. Rodent outbreaks in Europe: Dynamics and damage. In *Rodent Outbreaks—Ecology and Impacts*; Singleton, G.R., Belmain, S., Brown, P.R., Hardy, B., Eds.; International Rice Research Institute: Los Baños, Philippines, 2010; pp. 207–223.
62. Jacob, J.; Manson, P.; Barfknecht, R.; Fredricks, T. Common vole (*Microtus arvalis*) ecology and management: Implications for risk assessment of plant protection products. *Pest Manag. Sci.* **2014**, *70*, 869–878. [[CrossRef](#)]
63. Reil, D.; Imholt, C.; Eccard, J.A.; Jacob, J. Beech fructification and bank vole population dynamics—combined analyses of promoters of human Puumala virus infections in Germany. *PLoS ONE* **2015**, *10*, e0134124. [[CrossRef](#)]
64. Tersago, K.; Verhagen, R.; Vapalahti, O.; Heyman, P.; Ducoffre, G.; Leirs, H. Hantavirus outbreak in Western Europe: Reservoir host infection dynamics related to human disease patterns. *Epidemiol. Infect.* **2011**, *139*, 381–390. [[CrossRef](#)]
65. Grinnell, J. *An Account of the Mammals and Birds of the Lower Colorado Valley, with Especial Reference to the Distributional Problems Presented*; University of California Press: Berkeley, CA, USA, 1914; Volume 12, pp. 51–294.
66. Meinig, H.U.; Boye, P.; Hutterer, R.; Dähne, M. Rote Liste und Gesamtartenliste der Säugetiere (Mammalia) Deutschlands. *Nat. Biol. Vielfalt* **2020**, *170*, 1–673. [[CrossRef](#)]
67. RCoreTeam. *R: A Language and Environment for Statistical Computing*; R Foundation for Statistical Computing: Vienna, Austria, 2020.
68. Brooks, M.E.; Kristensen, K.; van Benthem, K.J.; Magnusson, A.; Berg, C.W.; Nielsen, A.; Skaug, H.J.; Maechler, M.; Bolker, B.M. glmmTMB balances speed and flexibility among packages for zero-inflated generalized linear mixed modeling. *R J.* **2017**, *9*, 378–400. [[CrossRef](#)]
69. Akaike, H. A new look at the statistical model identification. *IEEE Trans. Autom. Control* **1974**, *19*, 716–723. [[CrossRef](#)]
70. Wickham, H. *Ggplot2: Elegant Graphics for Data Analysis*; Springer: New York, NY, USA, 2009; Volume 1, p. 3.
71. Braun, M.; Dieterlen, F. *Die Säugetiere Baden-Württembergs*, 2nd ed.; Ulmer Verlag: Stuttgart, Germany, 2005.
72. Niedzialkowska, M.; Konczak, J.; Czarnomska, S.; Jędrzejewska, B. Species diversity and abundance of small mammals in relation to forest productivity in northeast poland. *Ecoscience* **2010**, *17*, 109–119. [[CrossRef](#)]
73. Suchomel, J.; Purchart, L.; Cepelka, L. Structure and diversity of small-mammal communities of lowland forests in the rural central European landscape. *Eur. J. For. Res.* **2012**, *131*, 1933–1941. [[CrossRef](#)]
74. Bryja, J.; Rehak, Z. Community of small terrestrial mammals (Insectivora, Rodentia) in dominant habitats of the Protected Landscape Area of Poodri (Czech Republic). *Folia Zool.* **1998**, *47*, 249–260.
75. Heroldova, M.; Bryja, J.; Zejda, J.; Tkadlec, E. Structure and diversity of small mammal communities in agriculture landscape. *Agric. Ecosyst. Environ.* **2007**, *120*, 206–210. [[CrossRef](#)]
76. Pearce, J.L.; Schuurman, D.; Barber, K.N.; Larrivee, M.; Venier, L.A.; McKee, J.; McKenney, D. Pitfall trap designs to maximize invertebrate captures and minimize captures of nontarget vertebrates. *Can. Entomol.* **2005**, *137*, 233–250. [[CrossRef](#)]
77. Weary, B.P.; Mendez, P.K.; Lew, S.E.; Nyman, B.L.; Will, K.W. Performance of ramp and pitfall traps for arthropods and non-target vertebrates in californian oak woodland and chaparral habitats. *Pan-Pac. Entomol.* **2019**, *95*, 21–32. [[CrossRef](#)]
78. McLeod, M.A.; Koronkiewicz, T.J. Comparison of capture and escape rates between different types of portable cowbird traps. *Wildl. Soc. Bull.* **2014**, *38*, 611–618. [[CrossRef](#)]
79. Cha, D.H.; Hesler, S.P.; Park, S.; Adams, T.B.; Zack, R.S.; Rogg, H.; Loeb, G.M.; Landolt, P.J. Simpler is better: Fewer non-target insects trapped with a four-component chemical lure vs. a chemically more complex food-type bait for *Drosophila suzukii*. *Entomol. Exp. Appl.* **2015**, *154*, 251–260. [[CrossRef](#)]
80. Campbell, S.; Cook, S.; Mortimer, L.; Palmer, G.; Sinclair, R.; Woolnough, A.P. To catch a starling: Testing the effectiveness of different trap and lure types. *Wildl. Res.* **2012**, *39*, 183–191. [[CrossRef](#)]
81. Anthony, N.M.; Ribic, C.A.; Bautz, R.; Garland, T. Comparative effectiveness of longworth and sherman live traps. *Wildl. Soc. Bull.* **2005**, *33*, 1018–1026. [[CrossRef](#)]
82. Von Blanckenhagen, F.; Städler, T. Small mammal communities in agricultural landscapes in Germany: Review of field data over the last decade. *Jul. -Kühn-Arch.* **2011**, *432*, 75. [[CrossRef](#)]
83. Boyard, C.; Vourc’h, G.; Barnouin, J. The relationships between *Ixodes ricinus* and small mammal species at the woodland-pasture interface. *Exp. Appl. Acarol.* **2008**, *44*, 61–76. [[CrossRef](#)] [[PubMed](#)]
84. DWD. *Nationaler Klimareport*, 4th ed.; Deutscher Wetterdienst (DWD): Potsdam, Germany, 2020.
85. Eccard, J.A.; Ylönen, H. Initiation of breeding after winter in bank voles: Effects of food and population density. *Can. J. Zool.* **2001**, *79*, 1743–1753. [[CrossRef](#)]
86. Jacob, J. Body weight dynamics of common voles in agro-ecosystems. *Mammalia* **2003**, *67*, 559–566. [[CrossRef](#)]
87. Tkadlec, E.; Zejda, J. Density-dependent life histories in female bank voles from fluctuating populations. *J. Anim. Ecol.* **1998**, *67*, 863–873. [[CrossRef](#)] [[PubMed](#)]

88. Gurnell, J.; Little, J. The influence of trap residual odour on catching woodland rodents. *Anim. Behav.* **1992**, *43*, 623–632. [[CrossRef](#)]
89. Brouard, M.J.; Coulson, T.; Newman, C.; Macdonald, D.W.; Buesching, C.D. Analysis on population level reveals trappability of wild rodents is determined by previous trap occupant. *PLoS ONE* **2015**, *10*, e0145006. [[CrossRef](#)]
90. Jacob, J.; Brown, J.S. Microhabitat use, giving-up densities and temporal activity as short and long term anti-predator behaviors in common voles. *Oikos* **2000**, *91*, 131–138. [[CrossRef](#)]
91. Yletyinen, S.; Norrdahl, K. Habitat use of field voles (*Microtus agrestis*) in wide and narrow buffer zones. *Agric. Ecosyst. Environ.* **2008**, *123*, 194–200. [[CrossRef](#)]
92. Clapperton, B.K. *A Review of the Current Knowledge of Rodent Behaviour in Relation to Control Devices*; Science & Technical Publication: Wellington, New Zealand, 2006; Volume 263, pp. 1–55.
93. Kok, A.D.; Parker, D.M.; Barker, N.P. Rules of attraction: The role of bait in small mammal sampling at high altitude in South Africa. *Afr. Zool.* **2013**, *48*, 84–95. [[CrossRef](#)]
94. Hansen, S.C.; Stolter, C.; Jacob, J. Effect of plant secondary metabolites on feeding behavior of microtine and arvicoline rodent species. *J. Pest Sci.* **2016**, *89*, 955–963. [[CrossRef](#)]